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Russian Consensus on "Hyperammonemia in Adults": The 2021 Version (in English)

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Summary

Hyperammonemia is an acute or chronic intoxication with ammonia and ammonium associated with elevated ammonia levels in serum due to either its increased production and/or decreased detoxification. Hyperammonemia can result from a variety of causes and clinically presents with unspecific signs and symptoms, including asthenia, encephalopathy, liver steatosis or fibrosis, and sarcopenia. With impaired liver function, hyperammonemia most frequently manifests in (micro)encephalopathy. Thus in case of unexpect change in mental status hyperammonemia must be excluded as fast as possible. An express method of photometric assay is informative enough to determine the ammonia levels. The following hyperammonemia classification is proposed: a) by ammonia levels (normal level: $\leq 60 \, \mu$ mol/L; mild (Grade 1): $\leq 100 \, \mu$ mol/L; moderate (Grade 2): $\leq 200 \, \mu$ mol/L; and severe (Grade 3): $> 200 \, \mu$ mol/L); b) by etiopathogenesis (hereditary (congenital), functional (physiological), acquired (hepatic, extrahepatic, mixed)); c) by clinical presentation (transient, recurrent or persistent, constant (stable, without treatment), covert). Treatment for hyperammonemia is aimed at treating the primary disease and includes a diet that is restricted in animal protein but contains sufficient vegetable protein, limited physical activities, and use of intestinal non-absorbable antibiotics (rifaximin-alpha) as well as pre- and probiotics. L-ornithine-L-aspartate (LOLA) is a baseline therapeutic product administered in a number of scenarios to correct the level of hyperammonemia.

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Резюме

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Гипераммонемия — это острая или хроническая интоксикация аммиаком и аммонием, связанная с повышенным уровнем аммиака в сыворотке крови из-за его повышенной выработки и / или снижения детоксикации. Гипераммонемия может быть результатом множества причин и клинически проявляться неспецифическими признаками и симптомами, включая астению, энцефалопатию, стеатоз или фиброз печени и саркопению. При нарушении функции печени гипераммониемия чаще всего проявляется (микро) энцефалопатией. Таким образом, в случае неожиданного изменения психического статуса необходимо как можно быстрее исключить гипераммониемию. Экспресс-метод фотометрического анализа достаточно информативен для определения уровня аммиака. Предлагается следующая классификация гипераммонемии: а) по уровням аммиака (нормальный уровень: ≤ 60 мкмоль / л; легкая (степень 1): \leq 100 мкмоль / л; средняя (степень 2): \leq 200 мкмоль / л; и тяжелая (степень 3):> 200 мкмоль / л); б) по этиопатогенезу (наследственный (врожденный), функциональный (физиологический), приобретенный (печеночный, внепеченочный, смешанный)); в) по клинической картине (преходящая, рецидивирующая или стойкая, постоянная (стабильная, без лечения), скрытая). Лечение гипераммониемии направлено на лечение основного заболевания и включает диету, ограниченную животным белком, но содержащую достаточное количество растительного белка, ограниченную физическую активность и использование кишечных невсасывающихся антибиотиков (рифаксимин-альфа), а также пре- и пробиотиков. . . . L-орнитин-L-аспартат (LOLA) — это базовый терапевтический продукт, применяемый в ряде сценариев для коррекции уровня гипераммониемии.

Ключевые слова: гипераммониемия, азот, аммиак, аммоний, L-орнитин L-аспарагинат, рифаксимин-альфа, пробиотики, пребиотики, лактитол, лактулоза, минимальная печеночная энцефалопатия, фиброз печени

Конфликт интересов. Авторы заявляют об отсутствии конфликта интересов.

Justification

Given the large number of reports on the peculiarities of liver lesions related to severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection [1], a team of experts who participated in the 23rd Congress of the Scientific Society of Gastroenterologists of

Russia and the 15th National Congress of Therapists on November 19, 2020, decided to make additions to the Russian Consensus, "Hyperammonemia in Adults," published in early 2020 [2, 3].

1. Provision 1

Interconverted biologically active molecules (gaseous ammonia NH, and ammonium cation NH, are

end-products of protein catabolism that are produced by biological tissues and urease-producing bacteria.

1.1. Comments:

Ammonia is mainly metabolized in liver cells and muscle tissue via the synthesis of urea and glutamine, which are excreted from the body through urine, feces, and exhaled air.

The main mechanisms of ammonia formation in the human body are as follows:

- 1. Non-oxidative deamination of certain amino acids (serine, threonine, and histidine) in the liver
- 2. Oxidative deamination of glutamic acid in all tissues (except muscle), especially in the liver and kidneys
- 3. Deamination of glutamic and aspartic acid amides in the liver and kidneys
- 4. Catabolism of biogenic amines in all tissues, but mostly in the nervous tissue
- 5. Hydrolytic deamination in intensively working muscles
- 6. Breakdown of glutamine the main source of energy for the cells of the small intestinal mucosa
- 7. Degradation of purine and pyrimidine bases in all tissues
- 8. Urease-producing microorganism activity in the stomach, colon, and urinary tract

A significant amount of ammonia is formed as a by-product of the metabolism of intestinal bacteria in the colon, where ammonia enters the circulation via the portal vein. Customarily, the liver rapidly metabolizes ammonia from the blood of the portal vein and detoxifies it, such that the blood that is exported from the liver is practically free of ammonia [4].

At physiological pH values, up to 99% of ammonia is easily converted into ammonium ions, which are unable to pass through biological membranes and therefore remain inside the cell.

Ammonia is a toxic gas and its blood concentration in healthy individuals is relatively low (25–40 $\mu mol/L$). There are only trace amounts of free ammonia in the blood – no more than 1% of the compound circulates in the free form in the liquid component of blood.

There is no internationally standardized normal range for the level of ammonia in the blood; therefore, it depends on the technique and reagents used in the laboratory. The acceptable level of ammonia in the blood (normal ammonemia) is usually no more than 60 μ mol/L [5, 6].

Symptoms of chronic intoxication are observed when the ammonia level is 2- to 3-fold higher than the acceptable level; however, even a slight increase (by 30–50%) has an adverse effect on the body, mostly on the central nervous system in the form of headache, fatigue, and drowsiness (everyday fatigue, occupational fatigue, nervous or physical overexertion, etc.).

Nearly all ammonia is excreted from the body via the kidneys through urine in the form of urea, which is synthesized in the liver, while ammonium ion salts are formed in the tubular epithelial cells. Ammonia enters the hepatic and renal cells via the transport processes of non-toxic compounds (glutamine, asparagine, glutamic acid, and alanine) formed during the neutralization (binding) of ammonia [7]. Urease synthesis is the main method of neutralizing ammonia. Urea accounts for 80–85% of all nitrogen excreted from the body.

The amount of excreted urea depends on the protein intake from food. If the daily protein intake is 80–100 g, 25–30 g of urea is produced and excreted per day. Urea production occurs via the ornithine cycle (Krebs–Henseleit cycle) in the liver. This cycle has two functions: (1) the conversion of amino acid nitrogen into urea, which prevents the accumulation of toxic products, mainly ammonia, and (2) the synthesis of arginine and its replenishment in the body. The cycle involves two amino acids that are not protein constituents – ornithine and citrulline – and two proteinogenic amino acids – arginine and aspartic acid. The process includes five reactions: the first two occur in the mitochondria while the rest occur in the cytosol of hepatocytes.

Glutamate dehydrogenase and glutamine synthetase enzymes have regulatory functions and determine the speed of ammonia formation and neutralization. Some enzymes involved in urea production are present in the brain, red blood cells, and cardiac muscle; however, the whole set of enzymes is only present in the liver.

To synthesize 1 mole of urea via the ornithine cycle in the liver, 1 mole of ammonia and 1 mole of aspartic acid are required; thus, 6.3 g of ammonia and 50 g of aspartic acid are required for the daily synthesis of 25 g of urea.

1.2. Excreted urea consists of "ammonia" nitrogen and aspartic acid nitrogen in equal parts

If the detoxifying function of the liver is reduced, the conversion of ammonia into urea is hindered and alternative metabolic pathways for this purpose are activated in the muscular system, astrocytes of the central nervous system, and kidneys [8–10]. Skeletal muscle tissue and nervous tissue are the main ammonia detoxifiers [11].

The conversion of glutamate into glutamine begins in the astrocytes of the brain, where it is catalyzed by glutamine synthetase. In persistent hyperammonemia, in addition to elevated glutamine synthetase, the activity of other enzymes – in particular the glutamate transport protein [12]—is changed. This results in an increased glutamate concentration in the extracellular space, which in turn leads to cytotoxicity through the activation of excitatory neurotransmitters (excitotoxicity) [13].

A stable elevated level of nitrogen compounds leads to the excessive formation of free radicals and further dysfunction of astrocytes. Prolonged hyperammonemia may be accompanied by the production of defective neurotransmitters, leading to a change in the activity of the serotonin and glutamatergic systems, with an increase in the activity of the GABAergic system [14].

The hypothalamic center is the most sensitive area to hyperammonemia in the brain [13], which manifests as persistent anorexia and protein–energy deficiency, aggravating catabolic processes.

Myostatin, produced by myocytes, not only largely inhibits the growth of muscle mass, but also leads to its decrease [15], which in hyperammonemia in patients with liver cirrhosis clinically manifests as sarcopenia [15, 16]. Moreover, a correlation between

protein malnutrition and the risk of sepsis has been revealed [12].

Hyperammonemia induces autophagy, where damaged proteins are broken down but not processed [17, 18]. The presence of sarcopenia is one of

the prognostic factors of death in patients with liver cirrhosis (low Model for End-Stage Liver Disease [MELD] scores – a tool for scoring terminal stages of liver disease and predicting preimplantation survival) [11, 19].

2. Provision 2

There are various methods for determining the normal concentration of ammonia (NH_3) and ammonium (NH_4^+) in the body, the most common of which are the enzymelinked immunoassay (ELISA) and express methods.

2.1. Comments:

Ammonia (NH_3) is a gas; however, at a physiological pH, 97% of the ammonia that is present in the blood is in the form of NH_4^+ .

Historically, the direct enzymatic method has been the most commonly used method for determining ammonia levels (ammonemia). It involves the reaction of ammonium in the sample with α -oxoglutarate in the presence of glutamate dehydrogenase, which reduces nicotinamide adenine dinucleotide phosphate (NADPH) to form glutamate, NADP+, and water. Ammonia levels vary in venous, capillary, and arterial blood, although it normally occurs in the range of 11–50 μ mol/L[20, 21]. Although arterial blood ammonia is considered a more reliable indicator of systemic ammonia levels, venous blood ammonia is usually analyzed, which somewhat differs from arterial ammonia [22, 23].

It is important to note that the ammonia level in a collected blood sample may spontaneously increase due to a needlestick injury of the vein, hemolysis of red blood cells, or other causes [24, 25]. Stress, exercise, psychological fatigue, smoking, malnutrition, and alcohol abuse may also affect systemic ammonia levels.

The measurement of ammonia levels in the blood is mainly limited by the requirement to strictly ensure proper collection, processing, and delivery of a blood sample. The sample obtained for transportation to the laboratory should be stored at a temperature below 0 °C, and the plasma should be separated from the red blood cells within 15 min after collection. After separation of the plasma, ammonia remains stable for 4 h at 4 °C. These stringent conditions for analysis require the blood to be collected close to the clinical laboratory [26].

Given the high risk of false results, it is recommended to repeat the analysis at the same time and under the same conditions to confirm the result. There are

The express method is a photometric assay of ammonia in capillary blood using a PocketChem BA PA-4140 portable analyzer (Arkray Global Business, Inc., Kyoto, Japan). It is informative and easy to use.

different methods for determining ammonia in red blood cells, exhaled air, saliva, sweat, and urine [27].

The measurement of ammonia levels in exhaled air is a non-invasive, easy-to-use, and comfortable diagnostic method suitable for all patient groups. No processing of exhaled air is required, as opposed to many tests performed using serum or urine samples; however, there is no reliable evidence that ammonia levels in exhaled air correlate with those in arterial blood [28–30].

Ammonia levels may be determined using liver tissue in cases where a patient requires a diagnostic liver biopsy. It should be considered that the colorimetric kits for the determination of ammonia in tissue samples are rather expensive and sensitive to other sources of ammonia in the environment; additionally, these samples require work in a special cabinet or negative air pressure area, which is not always available at all laboratories. Moreover, long-term storage of a tissue sample, even at a temperature of –80 °C, may affect its stability and lead to false results that are lower than expected.

In 2018, a portable express blood ammonia analyzer, the PocketChem BA PA-4140 (Arkray Global Business, Inc., Kyoto, Japan), was registered in Russia. Its method of determination is photometric - based on microdiffusion - that is, indirect; it takes approximately 200 s to determine the ammonia concentration using this device. Capillary blood from a finger is used for the analysis; however, blood collection should be performed under the conditions described in the instructions. Fresh blood samples should be used. In this device, ammonium is converted to ammonia after passing through a semi-permeable membrane; this changes the color of the indicator, the wavelength of which is spectroscopically analyzed. The resulting concentration is automatically indicated. Calibration of the device and correction of the results are also carried out automatically.

3. Provision 3

Hyperammonemia can be classified as follows:

- Level of ammonia (in capillary blood detected using the express method):
 - ► Normal (physiological) level: ≤ 60 μmol/L
 - ➤ Hyperammonemia:
 - ⊳ Mild (Grade 1): ≤ 100 μmol/L
 - ⊳ Moderate (Grade 2): ≤ 200 μmol/L
 - ⊳ Severe (Grade 3): > 200 μmol/L

- Etiopathogenesis:
 - ➤ Hereditary (congenital)
 - ➤ Functional (physiological)
 - ➤ Acquired (hepatic, extrahepatic, mixed)
- Clinical presentation:
 - ▶ Transient
 - ➤ Recurrent or persistent
 - Constant (stable, without treatment)
 - ▶ Covert

3.1. Comments:

- 1. The team of experts believes that with an increase in the number of observations, it will be possible to clarify the presented classification of hyperammonemia.
- 2. Traditionally, there are two main types of hyperammonemia in clinical practice [31]:
 - Hereditary hyperammonemia, as a result of various genetic defects in the urea cycle enzymes.
 - Acquired hyperammonemia, due to a decrease in the activity of the ornithine cycle and glutamine synthetase reaction; most often hepatic – in liver cell failure, as well as after portosystemic shunt surgery with the development and progression of portal hypertension – or extrahepatic (non-cirrhotic).

To date, the following causes of non-cirrhotic hyperammonemia have been identified [32, 33]:

- A. Conditions associated with increased ammonia production
 - Infections with urease producers: Proteus mirablis, Klebsiella species, Escherichia coli, Morganella morganii, Providencia rettgeri, Corynebacterium (causative agents of diphtheria), Mycobacterium genavense, Herpes simplex [34], and Helicobacter pylori (a greater incidence of non-alcoholic fatty liver disease [NAFLD] has been reported in patients with helicobacteriosis [35, 36].
 - Hemato-oncological disorders and their treatment: multiple myeloma, chemotherapy for acute leukemia, bone marrow transplantation, and 5-fluorouracil [37].
 - Organ transplantation [37].
 - Protein load and increased catabolism: intensive physical exercise, seizures, long-term malnutrition or severe injury, parenteral nutrition, gas-

- trointestinal bleeding, steroid use, and bariatric surgery.
- Overexertion, circadian rhythm disorders, and nightshift work without sufficient rest, i.e., sleep deprivation [38].
- Hyperammonemic encephalopathy cases have been reported after Roux-en-Y gastric bypass (RYGB) surgery, the most common weight loss procedure performed in the USA [39–41]; a mortality rate approximating 50% has been reported in women aged 34–69 years without a history of liver disease [40–52]. The risk of a fatal outcome may be owing to the bariatric surgery-related hyperammonemia not being accompanied by any biochemical changes in hepatic metabolism.
- Hyperammonemia may also occur in women during pregnancy and childbirth; it is associated with impaired hydrolytic deamination in intensively working muscles and with eclampsia.
- In adults, partial enzyme deficiency may manifest as stress-associated diseases such as postpartum stress, acute intestinal infection, short bowel syndrome, and gastrointestinal bleeding or occur with heart and lung transplantation or parenteral nutrition with high nitrogen intake [43, 53, 54].
- B. Conditions associated with decreased ammonia production
 - Ureterosigmoidostomy.
 - Portal systemic shunts, including congenital.
 - Use of the following medicines: valproic acid, glycine, carbamazepine, ribavirin, sulfadiazine, pyrimethamine, and salicylates.
 - Congenital metabolic defects: ornithine cycle disorders, defects in the β -oxidation of fatty acids and organic acids, and impaired metabolism of pyruvate.

4. Provision 4

The determination of ammonia and ammonium levels in the liver has following purposes:

- 1. To identify the level of hyperammonemia and monitor the efficacy of therapy in patients with any adjudicated acute or chronic liver disease.
- 2. To identify hyperammonemia in patients with chronic exo- or endogenous intoxication as part of the diagnosis.
- 3. To identify hyperammonemia in patients with symptoms of encephalopathy of various grades, asthenia, chronic fatigue, and neuroses as part of the diagnosis.
- 4. To identify congenital defects of nitrogen metabolism in the pediatric population with psychological defects in personality development.
- To evaluate the rate of recovery of metabolism in stressful situations, such as physical and psychological stress, surgical interventions, and nutritional distress.
- For the purpose of other situations, as per a physician's recommendation.

5. Provision 5

Physiological (functional) hyperammonemia may occur in the following cases: postprandial (high-protein diet), after physical exertion (sports), after psychogenic overload (post-stress), fatigue (long-term sleep deprivation), and in other situations (e.g., pregnancy).

5.1. Comments:

Physiological hyperammonemia is a sign of metabolic disorders in the muscle tissue and often manifests as fatigue due to impaired adenosine triphosphate (ATP) synthesis. Increased adenylate cyclase activity in the case of impaired ATP resynthesis increases the produc-

tion of ammonium ions, shifting metabolism towards excessive lactate formation, the development of acidosis, and hyperventilation, that is, hyperpnea [55, 56].

A high-protein diet, malnutrition, overeating, intense physical exertion (mainly in men and bodybuilders),

total parenteral nutrition, and childbirth also causes an increase in ammonium in the body. Ammonia levels increase especially in cases when a high-protein diet is followed after malnutrition [57].

During anaerobic exercise, ammonia metabolizes to urea in the hepatocytes and skeletal muscles; muscle

fatigue occurs as a result of exercise and leads to a noticeable decrease in performance. The main factors in the development of physical and psychological fatigue are decreased muscle contractions, the accumulation of metabolic products of energy sources, and an imbalance of the internal environment.

6. Provision 6

Congenital hyperammonemia is caused by a deficiency or defect in Krebs cycle enzymes – carbamoyl phosphate synthetase (hyperammonemia type 1), ornithine carbamoyltransferase (hyperammonemia type 2),

citrullinase (citrullinemia), argininosuccinate lyase deficiency (argininosuccinaturia), or arginase (hyperargininemia)—and is observed, as a rule, in early childhood.

6.1. Comments:

There are several types of genetic diseases caused by a deficiency or defect in an enzyme:

- Hyperammonemia type 1: based on a defect in carbamoyl phosphate synthetase I
- Hyperammonemia type 2: based on a defect in ornithine carbamoyltransferase
- Citrullinemia: based on a defect in argininosuccinate synthetase
- Argininosuccinaturia: based on a defect in argininosuccinate lyase
- · Hyperargininemia: based on an arginase deficiency

Some orphan diseases have also been reported in recent years. The rare clinical syndrome of hyperornithinemia–hyperammonemia–homocitrullinuria (HHH), ORPHA:415, Online Mendelian Inheritance in Man (OMIM) number 238970, have been described, which is a genetic disorder of the urea cycle caused by mutations in the *LC25A15* gene (MIM 603861, alternatively named *ORNT1*) encoding mitochondrial ornithine carrier ORC1. HHH syndrome is a heterogeneous

disease with various symptoms ranging from mild to severe, regardless of the age of the child. Symptoms include learning difficulties, mild neurogenic disorders to coma, lethargy, liver failure, and seizures [58].

Hyperinsulinism/hypoglycemia (HI/HA) syndrome has also been described; it has the second highest incidence after congenital hyperinsulinism, and is caused by mutations in the mitochondrial enzyme – glutamate dehydrogenase – which regulates the GLUT1 glucose transmitter. Clinically, HI/HA syndrome manifests as episodes of postprandial protein- and leucine-sensitive hypoglycemia with persistent hyperammonemia [59].

Furthermore, there are reported cases of hyperammonemia associated with genetic lysinuric protein intolerance caused by mutations in the *SLC7A7* gene, which determines the membrane transport of dibasic amino acids (ornithine, arginine, and lysine). In the absence of protein intake through food, the ammonia level is within the normal range; however, it increases significantly after the consumption of proteins, which can lead to coma [60].

7. Provision 7

Acquired hyperammonemia can be divided into the following types according to the pathogenesis:

- Hepatogenic: infectious, toxic, infectious-toxic, and metabolic
- Gastroenterogenic
- Urogenic
- · Vascular: for example, in portal hypertension
- Postoperative, including post-transplant
- Toxic: medicinal, including cytostatic, as well as post-radiation, oncological, oncohematological, associated with burns, smoking, and alcohol
 - Catabolic: high-protein diet, deficient diets, anaerobic exercise, age-related sarcopenia

7.1. Comments:

Acquired hyperammonemia may have an extrahepatic origin [33, 55, 61]. Toxically elevated ammonia levels have been reported in patients with hypovolemia, as well as in those without liver cirrhosis with bleeding from various parts of the gastrointestinal tract, heart failure, cor pulmonale, shunt surgeries, and some endocrine disorders (decompensated diabetes mellitus and severe thyrotoxicosis) [56].

Hyperammonemic encephalopathy is a rare complication of chemotherapy-resistant multiple myeloma [62], and has been described in leukemia due to catabolic processes [37] as well as in the acquired deficiency of enzymes of the ornithine cycle of urea synthesis, microvesicular steatosis of the liver (Reye's syndrome), liver perfusion disorder, metabolic alkalosis

and acidosis, bacterial overgrowth syndrome, and long-term constipation.

Hyperammonemia may be observed in any pathologies with increased protein catabolism, such as massive blood loss, severe burns, tissue compression or crush syndrome, extensive purulent necrosis, gangrene of the extremities, hyperthermia of various origins, and sepsis. These disorders cause loss of muscle mass, lack of antioxidant protection, and significantly suppressed immunity. An increase in ammonia, or imbalance in the ionized and non-ionized forms, is registered in autism; its role in the development of Alzheimer's disease is also not excluded. Viruses causing acute respiratory infections have been shown to decrease the activity of carbamoyl phosphate synthetase – the main

enzyme of the ornithine cycle – which in turn leads to the accumulation of the substrate of this enzyme and its precursors in the blood.

Moreover, hyperammonemia may be observed in pathologies of the lower urinary tract that cause difficulty in urine outflow complicated by infections caused by urease-producing bacteria (*Proteus* species, *Corynebacterium* species, *Klebsiella* species, *Morganella morganii*, etc.). In this case, the resulting free ammonia diffuses into the blood [63]. A correlation between small intestinal bacterial overgrowth (SIBO), capillary blood ammonia levels, and cognitive impairment has been established [64]. Hyperammonemia after transurethral resection of the prostate has been described [65].

Cases of hyperammonemia have also been registered after lung transplantation [66], with ureaplasma among the causes [67], as well as after kidney transplantation [68, 69].

Furthermore, an increase in blood ammonia level is associated with the administration of a number of medicinal products, including salicylates, tetracycline, glucocorticoids, asparaginase, 6-azauridin, allopurinol, thiazide diuretics, ethacrynic acid, isoniazid, and carbamazepine. [61]. Valproate-induced hyperammonemic encephalopathy in patients with epilepsy is not a rare complication and sometimes has a fatal outcome. In these cases, the increase in neurotoxin level is caused by the depletion of N-carbamylglutamate; it may develop rapidly and with long-term use of the medication [70]. The mechanism of drug-related hyperammonemia is not always clear. Secondary infections, hypovolemia, and a history of constipation are some of the risk factors involved in its development.

Hyperammonemia may also develop due to alcohol or substance abuse. One cigarette increases the level of ammonia in the blood by 10 μ mol/L.

8. Provision 8

The most researched clinical manifestations of hyperammonemia are hepatic encephalopathy (HE; including minimal and so-called "covert" encephalopathy, without observed clinical symptoms), subclinical hepatic failure due to endothelial dysfunction, and activation of hepatic stellate cells leading to aggravation of hepatic fibrogenesis.

8.1. Comments:

The Number Connection Test (NCT) may be used as a screening method for encephalopathy. HE is the most common manifestation of hyperammonemia.

There are several proposed theories and stages of HE pathogenesis:

- The binding of ammonia during glutamate synthesis causes depletion of α-ketoglutarate in the tricarboxylic acid cycle, with a decrease in ATP production and impaired cell activity.
- NH₄⁺ ions cause alkalization of blood plasma. Simultaneously, the affinity of hemoglobin for oxygen (the Bohr effect) increases; consequently, hemoglobin does not release oxygen in the capillaries, resulting in cellular hypoxia.
- The accumulation of free NH₄⁺ ions in the cytosol affects the membrane potential and activity of intracellular enzymes as it competes with Na⁺ and K⁺ pumps.
- Glutamine, the product resulting from the binding
 of ammonia to glutamic acid, is an osmotically active substance. This leads to water retention in and
 subsequent swelling of the cells, which causes tissue
 swelling. In the case of nervous tissue, this may cause
 cerebral edema, coma, and death.
- Glutamine is removed from the central nervous system via a transport system, by means of exchange; therefore, an increase in the rate of ammonia release is accompanied by an increase in the transport of aromatic amino acids from the blood. This causes inhibition of the enzymatic system, which normally converts tyrosine into dopamine and noradrenaline. Consequently, the initial compounds are metabolized by the formation of false neurotransmitters similar in structure to true adrenergic neurotransmitters which replace the latter at neuromuscular synapses, but with a 50-fold lower efficiency in transmitting nerve impulses.

- The accumulation of false neurotransmitters in the central nervous system (octopamine, phenylethylamine, tyramine, and phenylethanolamine) as well as serotonin – the metabolic product of tryptophan which is a neurotransmitter with predominantly inhibitory effects – contributes to depression of the nervous system, deterioration of brain functions, and development of encephalopathy.
- The accumulation of ammonia and mercaptans in the blood causes rapid progression of liver failure due to their pronounced inhibitory effect on hepatocyte proliferation.

The pathogenesis of HE is based on an imbalance in amino acids in the brain, leading to astroglial swelling and dysfunction, namely, altered postsynaptic receptors and neurotransmission, impaired permeability of the blood-brain barrier, and decreased energy supply to neurons. The severity of clinical manifestations often correlates with the level of ammonia in the blood serum. Depending on the severity of the brain disorder, four grades of HE are distinguished (*Table 1*).

In recent years, progress in diagnostic techniques has enabled distinguishing symptomatic HE stages (disorientation, ataxia, and coma) from the stage presenting with minimal clinical symptoms (covert HE).

Covert HE can be diagnosed using special questionnaires or the method of evoked potential. Often, patients and their relatives do not perceive observed cognitive and psychomotor disturbances, such as difficulties with decision-making or a reduction in psychomotor speed, as symptoms of the disease. It should be taken into account that clinical symptoms of covert HE include increased fatigue, weakness, irritability, sleep inversion (drowsiness during the day and insomnia at night), speech disorders, impaired handwriting, difficulty focusing while driving and performing work that requires increased focus, tremor, and hypoactive

Table 1. West Haven criteria for grading hepatic encephalopathy

Note:

HE: hepatic encephalopathy; NCT: Number Connection Test; EEG: electroencephalography

HE grade	Level of Consciousness	Flapping tremor (asterixis)	NCT time (s)	EEG, alpha rhythm frequency (waves/s)	Arterial ammonia level
0	No changes	No	15-30	8.5-12	Normal (11–55 μmol/L)
1	Sleep pattern disturbance, confusion, euphoria or anxiety, shortened attention span, difficulty performing arithmetic addition	Rare flapping tremor (1–2 movements/30 s) (asterixis)	31–50	7–8	Increased, up to 1.33-fold higher than the upper limit of normal
2	Lethargy or apathy, minimal disorientation of time and place, personality change, inappropriate behavior, difficulty performing arithmetic subtraction	Irregular tremor (3–4 movements/30 s)	51-80	5-7	Increased, 1.33–1.67-fold higher than the upper limit of normal
3	Somnolence but responsive to verbal stimuli, gross disorientation	Frequent tremor (5–10 movements/30 s)	81–120	3-5	Increased, 1.67– 2-fold higher than the upper limit of normal
4	Coma	Almost constant flapping tremor	> 120 (inability to finish the test)	< 3	Increased, > 2-fold higher than the upper limit of normal

Table 2.
Differential diagnosis of hepatic encephalopathy and conditions accompanied by cognitive impairment

Note:

Nosological autonomy of alcoholic dementia is not universally recognized owing to the pathogenic heterogeneity of cognitive disorders associated with alcohol abuse.

MRI: magnetic resonance imaging; HIV: human immunodeficiency virus; TSH: thyroid stimulating hormone

Diseases and syndromes	Causes	Clinical signs and neurovisualization data
Wernicke–Korsakoff syndrome	Thiamine deficiency-related encephalopathy. The main cause is alcohol abuse, as well as uncontrollable vomiting, malnutrition, and bariatric surgeries.	Acute or subacute onset, possible progression to coma, characteristic regress of symptoms with appropriately timed thiamine-replacement therapy. Classic triad (16–38% of cases): ocular disturbances (nystagmus, ocular disorders, abnormal papillary responses), ataxia, changes in consciousness/behavior. MRI findings: symmetric hyperintense lesions in mamillary bodies, thalami, or periaqueductal grey matter. Specificity of changes, 93% [74].
Marchiafava–Bignami disease	Corpus callosum primary degeneration (demyelination) in adults and older patients. Rare syndrome associated with abuse of poor-quality red wine.	Typical gradual onset with steady progression. Clinical signs are associated with cortical disorders: overt memory derangement, aphasia, apraxia, episodes of agitation followed by depression, propensity to violence, auditory and visual hallucinations. Subcortical disorders (athetosis, chorea, tic disorders), gait disturbances (irregular step length, absence of anteropulsion, shuffling "skier's" gait). MRI findings: hyperintense lesions in the corpus callosum on T2-weighted MRI in the acute phase, with cysts in necrotic areas [75, 76].
Alcoholic dementia*	Long-term alcohol abuse	Gradual onset and slow progression: impaired attention span, impaired short-term memory, thought flattening, primitive sense of humor, personality changes (anger, aggression). It differs from vascular dementia in having more pronounced delayed recall and recognition, and from Alzheimer's disease due to the absence of severe memory deficit. MRI findings: diffuse cerebral atrophy with more pronounced atrophy of the frontal cortex and cerebellum, characteristic dilatation of the third ventricle [75, 77].
Alcohol withdrawal syndrome	Complete withdrawal from alcohol use or its reduction in individuals with physical addiction	Characteristic generalized mild or moderate tremor, tachycardia, skin hyperemia, hypertension. Behavioral disturbances: overt anxiety, sleep disturbances (nightmares). In the worst-case scenario, development of acute psychosis (delirium tremens) with auditory, visual, and olfactory hallucinations. A specific pattern in neurovisualization has not been described [75].

Diseases and syndromes	Causes	Clinical signs and neurovisualization data
Pellagra	Lack of niacin (vitamin B ₃) in patients with chronic alcoholism or malnutrition (homeless people, patients with anorexia). Cases have been reported in HIV, Crohn's disease, carcinoid syndrome, patients undergoing dialysis, and during the administration of certain medicines (isoniazid, ethionamide, 6-mercaptopurine).	Classic pellagra triad: dermatitis, diarrhea, dementia. Dermatitis: symmetric erythema, pruritus, vesicular rash on non-covered parts of the body – extremities, face, around the neck (Casal collar). Neuropsychiatric signs vary from headaches and irritability to memory loss and psychosis. A specific pattern in neurovisualization has not been described [78].
Hypothyroidism	Underactive thyroid gland	Clinical signs: memory and concentration disturbances, general lack of interest, hypomimia, tiredness and somnolence. Possible development of seizures, vestibular ataxia (abnormal gait, nystagmus). Rare neurological signs: psychosis and severe impairment of consciousness. Based on several clinical observations, hypothyroidism may be considered a factor, aggravating the development of HE and possibly inducing hyperammonemia [79]. A specific pattern in neurovisualization has not been described. Diagnosis is based on TSH, T ₃ , and T ₄ levels, and etiology of the disease.

muscle reflexes. Such dysfunctions may lead to serious car accidents with severe consequences [71, 72]. In this regard, diagnosing covert HE is of great importance for workers in many fields, such as vehicle drivers and automated equipment operators.

Among all the grades of HE, covert HE is the most difficult to differentially diagnose because, despite satisfactory time and place orientation, there are changes in psychomotor or neuropsychological test results. Clinical examination at this stage may reveal some cognitive dysfunctions and/or behavioral disorders, including mild disorientation, euphoria or anxiety, decreased focusing ability, difficulty performing

arithmetic addition or subtraction, and a change in sleep pattern [73]. At this stage, HE should be differentiated from a number of conditions accompanied by cognitive impairment (*Table 2*).

Given that there is a correlation between the level of hyperammonemia and the grade of HE, which can be determined using the NCT, it is possible to implicitly assume the severity of hyperammonemia using the same NCT. The severity of encephalopathy is calculated based on the speed of the test performance (*Table 3*).

For the convenience of the physician and the patient, an NCT can be carried out online as well.

Time (s)	HE grade
< 40	0
40-60	Covert
61-90	1, 1-2
91–120	2
121-150	3
> 150	4

Table 3.
Determination of hyperammonemia severity based on the time taken to complete the Number Connection Test

Note:

HE: hepatic encephalopathy

9. Provision 9

Persistent and progressive hyperammonemia in NAFLD has a profibrogenic effect, contributing to

hepatic hemodynamic disorders, development of portal hypertension, and subsequently sarcopenia.

9.1. Comments:

In a study among patients with chronic liver disease with fibrosis grade 0–1 as per polyhepatography, portal hepatic hemodynamic disorders were reported. In patients with fibrosis grade 0, these changes were dynamic as per functional tests with deep breathing and nitrates. The obtained data demonstrate that intrahepatic microcirculation disorders at various locations already

develop in the early stages of chronic liver disease. Endothelial dysfunction was also observed in patients with chronic liver disease, including endothelium-dependent vasodilation impairment, an increase in nitric oxide metabolites in the peripheral blood, and impaired expression of nitric oxide synthases in the liver tissue (a decrease in the expression of endothelial

synthase and the expression of inducible nitric oxide synthase) [80, 81].

A British study [82] demonstrated *in vitro* and *in vivo* that hyperammonemia is the cause of:

- activation of stellate liver cells,
- reduction of cellular metabolism and proliferation of stellate liver cells,
- activation of the profibrogenic/pro-inflammatory profile of stellate cells,
- intrahepatic hemodynamic disorders and an increase in portal venous pressure,
- · stimulation of endoplasmic reticulum stress, and
- induction of formation of reactive oxygen species.

Experimental and clinical studies have demonstrated that in NAFLD, hyperammonemia develops at the precirrhotic stage owing to the decreased activity of enzymes of the ornithine cycle [83–88]. In mice with NAFLD that were fed a high-fat diet, a decrease in enzyme levels was observed; it was accompanied by a decrease in urea synthesis and an increase in the level of ammonia in the liver tissue in comparison with the control group. In patients with NAFLD and obesity, a significant decrease in the number and activity of enzymes of the ornithine cycle was observed as compared to healthy individuals, and these changes were much more pronounced at the stage of steatohepatitis than at the stage of steatosis [89–92].

10. Provision 10

Physiological (functional) transient, rapidly reversible, hyperammonemia does not require therapy. Pathogenetic therapy for hyperammonemia, regardless of the grade of severity, is aimed at treating the primary disease; this includes a diet that is restricted in animal protein and contains sufficient vegetable protein, limitation of physical exertion, and administration of intestinal non-absorbable antibiotics

10.1. Comments:

Modern treatment methods for hyperammonemia aim to reduce ammoniogenesis, absorb ammonia in the gastrointestinal tract, and activate ammonia removal—through the activation of ureagenesis—by treating the primary disease or supplementing intermediate products of the urea cycle and glutamine synthesis [93, 94].

A very low-protein diet may increase serum ammonia levels owing to the activation of muscle catabolism. In addition, limiting protein intake worsens nutritional status (stimulates protein catabolism in the body), which adversely affects the health of patients with HE [95]. In patients diagnosed with liver cirrhosis, the minimum daily dietary protein intake necessary to maintain a nitrogen balance is 0.8–1.0 g per kg of body weight [96]. Therefore, at present, some physicians recommend a normoprotein diet to patients with HE, while in cases of intolerance of vegetable protein substitution with animal protein is recommended [97–99].

LOLA, a stable salt of ornithine and aspartic acid, is an important substrate for the synthesis of glutamine and urea, the main components of deamination [100]. LOLA promotes the glutamine synthetase reaction in the liver and muscles. It also plays an important role in ensuring that aspartate enters the Krebs cycle; that is, it increases the synthesis of macroergic (high-energy) compounds and reduces the formation of lactic acid which, in turn, reduces the permeability of the blood-brain barrier to toxic substances.

The main pharmacological properties of LOLA include a double mechanism of action, as both amino acids enter the ornithine cycle, increasing protein tolerance and having an anabolic effect, increasing the energy potential of cells, and enhancing the utilization of lactic acid. The membrane-stabilizing effect provides the antioxidant effect of LOLA; this is especially important in chronic liver disease, primarily that with an alcoholic etiology.

(rifaximin-alpha), or pre- and probiotics (if indicated)

L-ornithine-L-aspartate (LOLA) is a baseline medicinal product, administered as monotherapy or in combination with other medicines. to help correct the level of hyperammonemia; it may potentiate the specified pharmacodynamic effects of medicines of other pharmacological classes.

L-ornithine:

- enters the urea cycle as a substrate (during citrulline synthesis),
- stimulates carbamoyl phosphate synthetase I (the first enzyme of the urea cycle),
- activates the glutamine synthetase reaction in the liver and muscles, and reduces the concentration of ammonia in the blood plasma,
- contributes to the normalization of the acid-base balance in the body,
- promotes the production of insulin and somatotropic hormone, and
- improves protein metabolism in diseases requiring parenteral nutrition.

L-aspartate:

- enters the urea cycle at the stage of synthesis of arginine succinate,
- is a substrate for the synthesis of glutamine,
- participates in the binding of ammonia in the perivenous blood, hepatocytes, brain, and other tissues,
- stimulates the synthesis of glutamine in muscles and perivenous hepatocytes,
- has a stimulating effect on inactive or affected liver cells.
- stimulates regeneration, and improves energy processes in affected liver tissue,
- participates in the cycle of tricarboxylic acids,
- is able to pass through cell membranes via active transport,
- participates in energy metabolism in the mitochondria, thus increasing the energy supply to tissues, and
- · has an anabolic effect on the muscles.

A meta-analysis of 10 randomized clinical trials including 884 patients reported that, in comparison

with a placebo, LOLA is significantly more effective in terms of improving mental status in HE (relative risk [RR]: 1.36, 95% confidence interval [CI]: 1.10–1.69, p = 0.005), including in overt HE (RR: 1.19, 95% CI: 1.01–1.39, overall efficacy: Z = 2.14, p = 0.03) and covert HE (RR: 2.15, 95% CI: 1.48–3.14, p < 0.0001). In this study, clinical improvement was accompanied by a significant decrease in the level of ammonia in the blood (mean difference: –17.50 μ mol/L, 95% CI: –27.73, –7.26, p = 0.0008); however these observations were less heterogeneous in clinical studies among the European population with less than 100 participants. Oral LOLA medications have demonstrated high efficacy in the treatment of covert HE [101].

Thus, the mechanism of action of LOLA suggests the appropriateness of using this medication for the treatment of patients with liver failure, particularly complicated HE, and precirrhotic and non-cyrhotic hyperammonemia. It is crucial to add LOLA to the therapy plan at the earliest stages of the disease. The duration of treatment depends on various factors, and it may be required over a long term. The use of LOLA in patients with liver failure and HE improves the function of hepatocytes as well as neurons [102].

In a double-blind, placebo-controlled study, Demure et al. examined the effect of L-ornithine on bicycle training tolerance, exhaustion rate, and ammonia metabolism during and after exercise in healthy volunteers. The concentration of ammonia in the blood plasma immediately and 15 min after additional exercises in the L-ornithine group was significantly lower than that in the placebo group. Thus, L-ornithine increased the ability to buffer ammonia both during and after exercise [103].

The hepatoprotective properties of LOLA have been observed in patients with chronic liver disease of

various etiologies [104–107]. Data from a multicenter, non-randomized, prospective cohort study among 1167 patients with chronic liver disease – including 648 patients with non-alcoholic steatohepatitis – demonstrated the efficacy and good tolerability of LOLA [107].

An increase in the concentration of ammonia is already recorded in patients with chronic liver disease at the precirrhotic stage [104]. In a study among patients with non-alcoholic fatty liver disease, it was shown that with the use of LOLA, the hyperammonemia that was initially present with fibrosis grade 0–1 decreased significantly and was accompanied by an improvement in the overall health status and laboratory test results. In another study employing rheohepatography, impaired portal blood flow was conclusively observed in hyperammonemia in patients with precirrhotic stages of chronic liver disease [108].

The treatment of 289 patients with non-alcoholic steatohepatitis using LOLA for 3 months, with good tolerability and high patient compliance, contributed to a decrease in ammonia levels, correlated with a decrease in vascular disorders, and lead to a significant improvement in clinical and biochemical parameters and quality of life [80]. LOLA improves intrahepatic blood flow in patients with various types of portal hemodynamic disorders. In addition, it reduces ammonia in the blood and, consequently, probably deactivates stellate liver cells, decreases their contractility, and improves hepatic blood flow.

LOLA also helps to prevent traffic accidents in drivers with chronic hepatitis C and minimal liver fibrosis [109]. Furthermore, hyperammonemia correction using LOLA potentiates the effects of the alpha- and beta-blocker, carvedilol, in patients with circulatory failure related to alcoholic liver disease [110].

Pharmacological methods of reducing ammonia and ammonium synthesis by intestinal microbiota

The main purpose of treating patients with hyperammonemia and HE is to inhibit the growth of urease-producing intestinal bacteria. The beneficial effect of rifaximin-alpha in the treatment of hyperammonemia has been demonstrated in several clinical studies [111–113].

Rifaximin alpha, a non-absorbable broad-spectrum antibiotic, is effective against most gram-positive and gram-negative aerobic and anaerobic bacteria. This medicinal product remains practically unabsorbed in the gastrointestinal tract; no more than 1% of the dose remains in the blood after oral administration under fasting conditions. Minimal absorption of the active substance into the blood plasma reduces the risk of systemic adverse effects and extraintestinal interactions with other medicinal products, and eliminates the need for dose adjustment in patients with liver disease.

It has been shown that rifaximin-alpha is more effective for the treatment of HE than non-absorbable disaccharides. Good Clinical Practice-compliant studies have confirmed that rifaximin-alpha may be more effective than lactulose in a number of patients with HE grades 1–3 [81, 114, 115]. The recommended total daily dose for adults (1200 mg/day) is usually divided into three doses of 400 mg each per day over 7–10 days; such a treatment course should be repeated monthly over an extended period, if necessary [116].

This medication has demonstrated clinical benefits in terms of lowering the level of ammonium in the blood and improving the clinical manifestations of HE. Several studies have also shown a decrease in the number of hospitalizations in patients with recurrent encephalopathy receiving rifaximin-alpha [117–120].

The use of probiotics enables competitive displacement of urease-producing pathogenic bacteria in the intestine. Studies that assessed the effect of probiotics on HE [121, 122] revealed a reduction in intestinal permeability and secretion of bacterial urease, an increase in the release of ammonia, and an improvement in the metabolic potential of the intestinal epithelium; moreover, probiotics play a role in reducing the concentration of ammonia in the portal blood given that they inhibit bacterial urease activity.

Since most probiotics produce acids that lower the intestinal pH, ammonia absorption decreases [123]. In addition, probiotics help to reduce inflammation and oxidative stress in liver cells, which results in an increase in the hepatic clearance of ammonia and a decrease in the absorption of other toxins.

A meta-analysis of 21 randomized clinical studies including 1420 patients with HE showed that probiotics reduce the clinical manifestations of HE, improve the quality of life of patients, and contribute to a decrease

in the concentration of ammonia in the blood plasma; however, they do not affect mortality rate [124].

Oral administration of *Lactobacillus* strains reduces blood ammonia levels in patients with liver cirrhosis [125]. Studies have shown that *Lactobacillus acidophilus* strains modify the intestinal flora and improve cognitive function in patients with liver cirrhosis, while the *Enterococcus faecium* SF68 strain increases protein tolerance, contributes to a decrease in ammonia levels, and improves mental state and psychometric indicators in the long-term treatment of patients with liver cirrhosis and HE grade 1–2 [126, 127]. Combinations of high-concentration probiotic strains (*Bifidobacterium sp., Lactobacillus ssp., and Streptococcus thermophilus*) have a significant effect

on the grade of hyperammonemia in patients with cirrhosis and HE [128].

Currently, probiotics are considered second- or third-line therapy for hyperammonemia and HE. Simultaneously, lactitol and lactulose prebiotics may be the treatment of choice in certain cases of HE [127, 129, 35].

Experts note that there are no proven correlations between the level of hyperammonemia and clinical signs of any diseases apart from liver disease. HE and liver fibrosis pathogenesis is widely studied and these conditions require a thoughtful and differential approach to treatment, which therefore should target numerous pathophysiologic factors that contribute to elevated levels of nitrogen metabolism end products.

11. Provision 11

Liver injuries related to coronavirus disease (COVID-19) have multiple etiologies and require pharmacological treatment. The disease prognosis is worsened by pre-existing chronic diseases, such as non-alcoholic fatty liver disease and alcoholic and toxic liver damage, including drug-induced and chronic viral hepatitis. The severity

criteria are an increase in cytolysis, ferritin level, hypoalbuminemia, and the presence of fibrosis.

Medication is required in these clinical cases to manage hyperammonemia, including the use of LOLA and monitoring levels of ammonia and ammonium in the peripheral blood.

11.1. Comments:

Frequent digestive system lesions caused by SARS-CoV-2, as well as its possible combination with gastrointestinal pathology, complicates the set of therapeutic and diagnostic measures for such patients and implies modification of the existing approaches and clinical guidelines.

The COVID-19 spike glycoprotein has a tropism for endothelial cells, hepatocytes, and cholangiocytes containing ACE2 receptors, which cause damage to parenchymal organs, including the liver and mucous membranes. Chronic liver disease aggravates the severity of the patient's condition.

Diarrhea, nausea, and vomiting may be among the first symptoms of COVID-19, in addition to the clinical signs of acute respiratory viral infection. Studies show rather heterogeneous data on the frequency of symptoms of digestive system lesions from the onset of the disease.

Fifty percent of patients with COVID-19 have a transient increase in transaminase activity, bilirubin levels (up to 2-fold higher than the upper limit of normal) caused by hyperactivation of the immune system, cytokine production, and inflammation (bystander hepatitis), together with viral damage to cholangiocytes and hepatotoxic effects secondary to the medications administered.

Patient examination should include:

- Clarification of digestive system complaints: anorexia, change in bowel habits, flatulence, nausea, vomiting, and abdominal pain
- Physical examination, determining the severity of the patient's condition, including: examination of the abdomen, palpation, percussion of the abdominal organs with the definition of areas of hyperactive bowel sounds, tenderness to deep palpation, and the

size of the liver and spleen according to Kurlov. It is also necessary to clarify the characteristics of the stool: frequency per day, volume, shape (consistency), color, smell, and pathological impurities

- Laboratory tests: alanine transaminase, alanine aminotransferase, gamma-glutamyl transpeptidase, alkaline phosphatase, and bilirubin
- Measurement of ammonia levels (for correction of hyperammonemia)
- Abdominal ultrasound (in case of clinical signs and laboratory findings suggestive of liver injury)

The disease prognosis is worsened by pre-existing chronic diseases, including:

- · non-alcoholic liver disease,
- alcoholic liver disease,
- · toxic liver damage,
- drug-induced liver injury, and
- viral hepatitis.

An increase in the level of ammonia in the peripheral blood has been reported in all hospitalized patients with COVID-19. It should be noted that a further increase in this value (above the cut-off point) results in severe disease progression with deterioration of the clinical state, which requires specialized respiratory support and the transfer of the patient to anesthesiology and resuscitation units. Thus, the grade of hyperammonemia may be considered a prognostic marker of disease severity [130].

Medication, LOLA, in these clinical cases is required to manage hyperammonemia while monitoring the levels of ammonia and ammonium in the peripheral blood [131].

12. Provision 12

Therapeutic indications for LOLA are:

- acute and chronic liver disease, accompanied by hyperammonemia,
- HE (covert and overt), and
- · steatosis and steatohepatitis (of various etiologies).

Depending on the severity of hyperammonemia, the recommended treatment courses are:

- Mild hyperammonemia (grade 1): LOLA 3.0-6.0 g (1-2 sachets) 3 times per day for 4 weeks; after the completion of treatment, performance of the NCT and measurement of the ammonia level in the blood
- Moderate hyperammonemia (grade 2): LOLA 3.0-6.0 g (1-2 sachets) 3 times per day for up to 3 months; during and after completion of treatment,

- performance of the NCT and measurement of the ammonia level in the blood
- Severe hyperammonemia (grade 3): LOLA 10.0–40.0 g (20.0–80.0 mL) per 400.0 mL of sodium chloride saline solution intravenously by slow drip infusion daily until clinical symptoms are resolved and ammonia levels in the blood are reduced. Thereafter, 3.0–6.0 g (1–2 sachets) 3 times per day for up to 3–6 months (if necessary, for a longer period); during and after completion of treatment, performance of the NCT and measurement of the ammonia level in the blood

If necessary, treatment courses may be repeated. Before using the medication, the patient should carefully read the instructions.

13. Conclusion

Hyperammonemia is a condition characterized by an elevated level of ammonia and ammonium in the blood. It can be physiological (transient, which does not require therapy) or pathological (related to the increased formation or decreased excretion of these toxic entities due to the reduced protective function of the liver). Chronic hyperammonemia manifests in the development of hepatic steatosis or fibrosis, encephalopathy, and sarcopenia. Hyperammonemia treatment involves a combination

of therapeutic options to neutralize and excrete toxic products of nitrogen metabolism in order to prevent life-threatening complications. Among the medications to treat hyperammonemia, L-ornithine-L-aspartate (LOLA) should be considered, which is available in a parenteral and oral dosage forms for the treatment of hepatogenic hyperammonemia and hepatic encephalopathy of different stages, and may help to mitigate the risk of liver disorder progression related to hyperammonemia.

References | Литература

- Pan L., Huang P., Xie X., Xu J., Guo D., Jiang Y. Metabolic associated fatty liver disease increases the severity of COVID-19: A meta-analysis. Dig Liver Dis. 2021 Feb;53(2):153–157. doi: 10.1016/j.dld.2020.09.007. Epub 2020 Sep 17. PMID: 33011088; PMCID: PMC7498251.
- Lazebnik L.B., Golovanova E. V., Alekseenko S. A., et al. Russian Consensus "Hyperammonemia in Adults". Experimental and Clinical Gastroenterology. 2019;(12):4– 23. (In Russ.). Doi: 10.31146/1682–8658-ecg-172–12– 4–23
- 3. Lazebnik L.B., Golovanova E.V., Alekseenko S.A., et al. Russian consensus "Hyperammonemia in adults". *Therapy.* 2020;4(38):36–51. (In Russ.). DOI: 10.18565/therapy.2020.4.36–51
- Murray R., Grenner D., Mayes P., Rodwell V. Human biochemistry. vol. 1. Moscow. BINOM. Laboratory of Knowledge, 2009. 384 p. (In Russ.).
- Kamyshnikov V.S. Clinical and laboratory diagnosis of liver diseases. Moscow. MEDpress-inform. 2013, 96 p. (In Russ.).
- Dolgov V. V., Menshikov V. V. Clinical laboratory diagnostics: National guidelines: vol. 1. Moscow. GEOTAR-Media, 2012, 928 p. (In Russ.).
- Nelson D. Leninger Principles of Biochemistry: in 3 vols. Moscow. BINOM. Laboratory of Knowledge Publ., 2014, 636 p. (In Russ.).
- 8. Pavlov C. S., Damulin I. V., Ivashkin V.T. Hepatic encephalopathy: pathogenesis, clinical presentation, diag-

- nostics, treatment. *Russian Journal of Gastroenterology, Hepatology, Coloproctology.* 2016;26(1):44–53. (In Russ.). https://doi.org/10.22416/1382–4376–2016–26–1–44–53.
- Zimmerman Ya. S. Hepatic encephalopathy: definition, etiology, pathogenesis factors, clinic, diagnostic and treatment methods. *Gastroenterology of St. Petersburg*. 2017, No. 2, pp. 68–75. (In Russ.).
- Nikonov E. L., Aksenov V. A. Hepatic encephalopathy. Russian journal of Evidence-based gastroenterology = Dokazatel'naya gastroenterologiya. 2017;6(4):25-31. (In Russ.). Doi: 10.17116/dokgastro20176425-31
- Ilchenko L. Y., Nikitin I. G. Hyperammonium in patients with precirrhosis stage: clinical reality? *The Russian Archives of Internal Medicine*. 2018;8(3):186–193.
 (In Russ.). https://doi.org/10.20514/2226-6704-2018-8-3-186-193
- Golovanova E. V. Ammonia as an actual problem of a doctor and patient in therapeutic practice. *Therapy*. 2018, no. 2, pp. 49–55. (In Russ.).
- Blachier F., Boutry C., Bos C., Tomé D. Metabolism and functions of L-glutamate in the epithelial cell of the small land large intestines. Am J Clin Nutr. 2009; 90: 814S-821S.
- 14. Newsholme E.A., Carrié A.L. Quantitative aspects of glucose and glutamine metabolism by intestinal cells. *Gut*. 1994;35(1 Suppl): S13-S17. doi: 10.1136/gut.35.1_suppl.s13
- Qiu J., Thapaliya S., Runkana A., et al. Hyperammonemia in cirrhosis induces transcriptional regulation of myo-

- statin by an NF-κB-mediated mechanism. *Proc Natl Acad Sci USA*. 2013 Nov 5;110(45):18162–7. doi: 10.1073/pnas.1317049110. Epub 2013 Oct 21. PMID: 24145431; PMCID: PMC3831479.
- Noiret L., Baigent S., Jalan R. Arterial Ammonia levels in cirrhosis are determined by systemic and hepatic hemodynamics, and by organ function: a quantitative modeling study. *Liver Int*. 2014 Jul;34(6): e45–55. doi: 10.1111/liv.12361. Epub 2013 Nov 20. PMID: 24134128.
- Qiu J., Tsien C., Samjhana T. Hyperammonemia-mediated autophagy in skeletal muscle contributes to sarcopenia of cirrhosis. Am J Physiol Endocrinol Metab. 2012 Oct 15;303(8): E983–93. doi: 10.1152/ajpendo.00183.2012. Epub 2012 Aug 14. PMID: 22895779; PMCID: PMC3469607.
- McDaniel J., Davuluri G., Hill EA., Moyer M., Runkana A., Prayson R., van Lunteren E., Dasarathy S.
 Hyperammonemia results in reduced muscle function independent of muscle mass. American Journal of Physiology Gastrointestinal and Liver Physiology Published. 2016; 310(3): G163-G170.
- Meza-Junco J., Montano-Loza A. J., Baracos V. E., Prado C. M., Bain V. G., Beaumont C., Esfandiari N., Lieffers J. R., Sawyer M. B. Sarcopenia as a prognostic index of nutritional status in concurrent cirrhosis and hepatocellular carcinoma. *J Clin Gastroenterol*. 2013 Nov-Dec;47(10):861–70. doi: 10.1097/MCG.0b013e318293a825. PMID: 23751844.
- Ong J.P., Aggarwal A., Krieger D., Easley K. A., Karafa M. T., Van Lente F., Arroliga A. C., Mullen K. D. Correlation between ammonia levels and the severity of hepatic encephalopathy. *American Journal of Medicine*. 2003 Feb 15;114(3):188–93. doi: 10.1016/s0002–9343(02)01477–8. PMID: 12637132.
- Mehmood M. A., Waseem T., Ahmad F. Z., Humayun M. A. Measuring Partial Pressure of Ammonia in Arterial or Venous Blood VS total Ammonia Levels in Hepatic Encephalopathy. J. Gastroenterol. Hepatol. 2013, Res. 2, pp. 602–606.
- Metz M. P. Ammonia, a roublesome analyte. Clinical Biochemistry. 2014 Jun;47(9):753. doi: 10.1016/j.clinbiochem.2014.05.044. Epub 2014 May 20. PMID: 24854702.
- 23. Adeva M.M., Souto G., Blanco N., Donapetry C. Ammonium metabolism in humans. *Metabolism-Clinical and Experimental*. 2012 Nov;61(11):1495–511. doi: 10.1016/j.metabol.2012.07.007. Epub 2012 Aug 24. PMID: 22921946.
- Maranda B., Cousineau J., Allard P., Lambert M. false positives in plasma ammonia measurement and their clinical impact in a pediatric population. *Clinical Biochemistry*. 2007 May;40(8):531–5. doi: 10.1016/j. clinbiochem.2007.01.024. Epub 2007 Mar 12. PMID: 17408610.
- Conway E. J. Apparatus for the micro-determination of certain volatile substances: The blood ammonia, with observations on normal human blood. *The Biochemical Journal*. 1935 Dec;29(12):2755–72. doi: 10.1042/ bj0292755. PMID: 16745964; PMCID: PMC1266825.
- Huizenga J.R., Gips C. H., Tangerman, A. The contribution of various organs to ammonia formation: A review of factors determining the arterial ammonia concentration.
 Annals of Clinical Biochemistry. 1996 Jan;33 (Pt 1):23–30. doi: 10.1177/000456329603300103. PMID: 8929062.
- Green A. When and how should we Measure Plasma Ammonia. *Annals of Clinical Biochemistry*. 1988 May;25 (Pt 3):199–209. doi: 10.1177/000456328802500301. PMID: 3041901.

- 28. Adrover R., Cocozzella D., Ridruejo E., Garcia A., Rome J., Podesta J. J. Breath-Ammonia Testing of Healthy Subjects and Patients with Cirrhosis. *Dig Dis Sci.* 2012 Jan;57(1):189–95. doi: 10.1007/s10620–011–1858–9. Epub 2011 Aug 13. PMID: 21842240.
- DuBois S., Eng, S., Bhattacharya R., Rulyak S., Hubbard T., Putnam D., Kearney D. J. Breath ammonia testing for diagnosis of hepatic encephalopathy. *Dig Dis Sci.* 2005; 50(10): 1780–4.
- Brannelly N. T., Hamilton-Shield J. P., Killard A. J. The Measurement of Ammonia in Human Breath and its Potential in Clinical Diagnostics. *Analytical Chemistry*, 2016, Vol. 46, pp. 490–501.
- 31. Vilstrup H., Amodio P., Bajaj J., Cordoba J., Ferenci P., Mullen K. D., Weissenborn K., Wong P. Hepatic encephalopathy in chronic liver disease: 2014 Practice Guideline by the American Association for the Study of Liver Diseases and the European Association for the Study of the Liver. *Hepatology*. 2014 Aug;60(2):715–35. doi: 10.1002/hep.27210. Epub 2014 Jul 8. PMID: 25042402.
- Yoshino M, Nishiyori J, Yamashita F, Kumashiro R, Abe H, Tanikawa K, Ohno T, Nakao K, Kaku N, Fukushima H. Ornithine transcarbamylase deficiency in male adolescence and adulthood. *Enzyme*. 1990;43(3):160–8. doi: 10.1159/000468724. PMID: 2095337.
- Laish I., Ben Ari Z. Noncirrhotic hyperammonaemic encephalopathy. *Liver Int*. 2011 Oct;31(9):1259–70. doi: 10.1111/j.1478–3231.2011.02550.x. Epub 2011 Jun 21. PMID: 21745294.
- Ghatak T, Azim A, Mahindra S, Ahmed A. Can Klebsiella sepsis lead to hyperammonemic encephalopathy with normal liver function? *J Anaesthesiol Clin Pharmacol*. 2013 Jul;29(3):415–6. doi: 10.4103/0970–9185.117079. PMID: 24106385; PMCID: PMC3788259.
- 35. Seung Joo Kang, Hwa Jung Kim, Donghee Kim, Aijaz Ahmed. Association between cagA negative Helicobacter pylori status and nonalcoholic fatty liver disease among adults in the United States. PLoS One. 2018 Aug 15;13(8): e0202325. doi: 10.1371/journal.pone.0202325. PMID: 30110395; PMCID: PMC6093702.
- Dogan Z., Filik L., Ergul B., Sarikaya M., Akbal E. Association between Helicobacter pylori and liver-to-spleen ratio: a randomized-controlled single-blind study. Eur. J. Gastroenterol. Hepatol. 2013; 25: 107–110. Doi: 10.1097/MEG.0b013e3283590c10
- Nott L., Price T. J., Pittman K., Patterson K., Fletcher J. Hyperammonemia encephalopathy: an important cause of neurological deterioration following chemotherapy. *Leuk Lymphoma*. 2007 Sep;48(9):1702–11. doi: 10.1080/10428190701509822. PMID: 17786705.
- 38. Plotnikova E. Yu., Vorosova O. A., Baranova E. N. et al. Night shift and hyperammonemia in doctors. *RMJ*. 2021;4:49–52. (In Russ.).
- 39. Samuel I., Mason E.E., Renquist K.E., Huang Y.H., Zimmerman M.B., Jamal M. Bariatric surgery trends: An 18 year report from the International Bariatric Surgery Registry. Am J Surg. 2006 Nov;192(5):657–62. doi: 10.1016/j.amjsurg.2006.07.006. PMID: 17071202.
- 40. Longitudinal Assessment of Bariatric Surgery (LABS) Consortium, Flum D. R., Belle S. H., King W. C., Wahed A. S., Berk P., Chapman W., Pories W., Courcoulas A., McCloskey C., Mitchell J., Patterson E., Pomp A., Staten M. A., Yanovski S. Z., Thirlby R., Wolfe B. Perioperative safety in the longitudinal assessment of bariatric surgery. N Engl J Med. 2009 Jul 30;361(5):445– 54. doi: 10.1056/NEJMoa0901836. PMID: 19641201; PMCID: PMC2854565.

- Limketkai B.N., Zucker S.D. Hyperammonemic Encephalopathy Caused by Carnitine Deficiency. *J Gen Intern Med.* 2008 Feb;23(2):210–3. doi: 10.1007/s11606–007–0473–0. Epub 2007 Dec 13. PMID: 18080167; PMCID: PMC2359173.
- Juhasz-Pocsine K, Rudnicki S. A., Archer R. L., Harik S. I. Neurologic complications of gastric bypass surgery for morbid obesity. *Neurology*. 2007 May 22;68(21):1843– 50. doi: 10.1212/01.wnl.0000262768.40174.33. PMID: 17515548.
- Summar M.L., Barr F., Dawling S., Smith W., Lee B., Singh R.H., Rhead W.J., Sniderman King L., Christman B.W. Unmasked adult-onset urea cycle disorders in the critical care setting. *Crit Care Clin*. 2005 Oct;21(4 Suppl): S1–8. doi: 10.1016/j.ccc.2005.05.002. PMID: 16227111.
- 44. Hu W.T., Kantarci O.H., Merritt J.L., 2nd, et al. Ornithine transcarbamylase deficiency presenting as encephalopathy during adulthood following bariatric surgery. *Arch Neurol.* 2007 Jan;64(1):126–8. doi: 10.1001/archneur.64.1.126. PMID: 17210820.
- Goodin K.M. PD, Platky K., Gowans G., et al. Asymptomatic carrier of Ornithine transcarbamylase deficiency unmasked by bariatric surgery. Annual Clinical Genetics Meeting: University of Louisville, Louisville, KY, 2010.
- Estrella J., Yee G., Wilcken B., Tchan M., Talbot M. Hyperammonemic encephalopathy complicating bariatric surgery: A case study and review of the literature. Surg Obes Relat Dis. 2014 May-Jun;10(3): e35–8. doi: 10.1016/j. soard.2013.10.020. Epub 2013 Nov 12. PMID: 24462307.
- Rogal S.S., Hu A., Bandi R., Shaikh O. Novel therapy for non-cirrhotic hyperammonemia due to a spontaneous splenorenal shunt. World J Gastroenterol. 2014 Jul 7;-20(25):8288–91. doi: 10.3748/wjg.v20.i25.8288. PMID: 25009405; PMCID: PMC4081705.
- Fenves A., Boland C. R., Lepe R., Rivera-Torres P., Spechler S. J. Fatal hyperammonemic encephalopathy after gastric bypass surgery. *Am J Med*. 2008 Jan;121(1): e1–2. doi: 10.1016/j.amjmed.2007.08.032. PMID: 18187055.
- 49. Acharya G., Mehra S., Patel R., Frunza-Stefan S., Kaur H. Fatal NonhepaticHyperammonemia in ICU Setting: A Rare but Serious Complication following Bariatric Surgery. Case Rep Crit Care. 2016;2016:8531591. doi: 10.1155/2016/8531591. Epub 2016 Apr 10. PMID: 27144037; PMCID: PMC4842030.
- 50. Kromas M.L., Mousa O.Y., John S. Hyperammonemia-induced encephalopathy: A rare devastating complication of bariatric surgery. *World J Hepatol.* 2015 May 8;7(7):1007–11. doi: 10.4254/wjh.v7.i7.1007. PMID: 25954483; PMCID: PMC4419094.
- 51. Fenves A.Z., Shchelochkov O.A., Mehta A. Hyperammonemic syndrome after Roux-en-Y gastric bypass. *Obesity* (Silver Spring). 2015 Apr;23(4):746–9. doi: 10.1002/oby.21037. Epub 2015 Mar 7. PMID: 25754921.
- Nagarur A., Fenves A.Z. Late presentation of fatal hyperammonemic encephalopathy after Roux-en-Y gastric bypass. *Proc (Bayl Univ Med Cent)*. 2017 Jan;30(1):41–43. doi: 10.1080/08998280.2017.11929521. PMID: 28127128; PMCID: PMC5242109.
- 53. Panlaqui O.M., Tran K., Johns A., McGill J., White H. Acute hyperammonemic encephalopathy in adult onset ornithine transcarbamylase deficiency. *Intensive Care Med.* 2008 Oct;34(10):1922–4. doi: 10.1007/s00134–008–1217–2. Epub 2008 Jul 24. PMID: 18651132.
- 54. Schultz R.E., Salo M.K. Under recognition of late onset ornithine transcarbamylase deficiency. *Arch Dis Child*.

- 2000 May;82(5):390–1. doi: 10.1136/adc.82.5.390. PMID: 10799432; PMCID: PMC1718304.
- Plotnikova E. Yu., Makarova M. P., Gracheva T. Yu. Possibilities of application of L-ornithine in sports medicine. Sports medicine: research and practice. 2016; 4: 28–35. (In Russ.).
- Hawkes N.D., Thomas G. A., Jurewicz A., Williams O. M., Hillier C. E., McQueen I.N., Shortland G. Non-hepatic hyperammonaemia: an important, potentially reversible cause of encephalopathy. *Postgrad Med J.* 2001 Nov;77(913):717–22. doi: 10.1136/pmj.77.913.717. PMID: 11677282; PMCID: PMCI742166.
- Welsh E., Kucera J., Perloff M. D. Iatrogenic hyperammonemia after anorexia. *Arch Intern Med.* 2010 Mar 8;170(5):486–8. doi: 10.1001/archinternmed.2009.549. PMID: 20212188.
- 58. Martinelli D., Diodato D., Ponzi E., et al. The hyperornithinemia–hyperammonemia-homocitrullinuria syndrome. *Orphanet J Rare Dis.* 2015 Mar 11;10:29. doi: 10.1186/s13023–015–0242–9. PMID: 25874378; PMCID: PMC4358699.
- 59. Mangla P., Hussain K., Ellard S., Flanagan S. E., Bhatia V. Diazoxide toxicity in a child with persistent hyperinsulinemic hypoglycemia of infancy: mixed hyperglycemic hyperosmolar coma and ketoacidosis. *J Pediatr Endocrinol Metab*. 2018 Aug 28;31(8):943–945. doi: 10.1515/jpem-2018-0112. PMID: 29958183.
- 60. Nunes V., Niinikoski H. Lysinuric Protein Intolerance. In: Adam M. P., Ardinger H. H., Pagon R. A., Wallace S. E., Bean L. J. H., Stephens K., Amemiya A., editors. GeneReviews* [Internet]. Seattle (WA): University of Washington, Seattle; 1993–2018. 2006 Dec 21 [updated 2018 Apr 12].
- Walker V. Severe hyperammonaemia in adult snot explained by liver disease. *Annals of Clin. Biochemistry*. 2012 May;49(Pt 3):214–28. doi: 10.1258/acb.2011.011206. Epub 2012 Feb 20. PMID: 22349554.
- 62. Lora-Tamayo J., Palom X., Sarra´ J., Gasch O., Isern V., Fernández de Sevilla A., Pujol R. Multiple myeloma and hyperammonemic encephalopathy: review of 27 cases. *Clin Lymphoma Myeloma*. 2008 Dec;8(6):363–9. doi: 10.3816/CLM.2008.n.054. PMID: 19064403.
- Kaveggia F. F., Thompson J. S., Schafer E. C., Fischer J. L., Taylor R. J. Hyperammonemic encephalopathy in urinary diversion with urea-splitting urinary tract infection. *Arch Intern Med.* 1990 Nov;150(11):2389–92. PMID: 2241451.
- 64. Vologzhanina L. G., Borodina E. N., Igumnova O. A., Trapeznikova A. A. Bacterial overgrowth syndrome with hyperammonemia as a cause of cognitive disorders. *Experimental and Clinical Gastroenterology*. 2021;(2):79–87. (In Russ.). doi:10.31146/1682-8658-ecg-186-2-79-87
- 65. Hoekstra P.T., Kahnoski R., McCamish M.A., Bergen W., Heetderks D. R. Transurethral Prostatic Resection Syndrome – A New Perspective: Encephalopathy with Associated Hyperammonemia. J. Urology. 1983 Oct;130(4):704–7. doi: 10.1016/s0022–5347(17)51414–7. PMID: 6887402.
- 66. Chen Catherine, Bain Karen B., Iuppa Jennifer A., et al. Hyperammonemia Syndrome After Lung Transplantation; A Single Center Experience. *Transplantation*. 2016 Mar;100(3):678–84. doi: 10.1097/TP.0000000000000868. PMID: 26335916.
- Matson K.M., Sonetti D.A. Successful treatment of Ureaplasma-induced hyperammonemia syndrome post-lung transplant. *Transpl Infect Dis.* 2019 Feb;21(1):

- e13022. doi: 10.1111/tid.13022. Epub 2018 Nov 22. PMID: 30403322
- 68. Nurmohamed S., Weenink A., Moeniralam H., Visser C. and Bemelman F. Hyperammonemia in Generalized Mycobacterium genavense Infection after Renal Transplantation. *Am. J. Transplantation*. 2007 Mar;7(3):722–3. doi: 10.1111/j.1600-6143.2006.01680.x. Epub 2007 Jan 22. PMID: 17250553.
- Li G.Z., Tio M. C., Pak L. M., et al. Noncirrhotic hyperammonemia after deceased donor kidney transplantation:
 A case report. Am J Transplant. 2019 Nov;19(11):3197–3201. doi: 10.1111/ajt.15545. Epub 2019 Sep 12. PMID: 31347272; PMCID: PMC6864227.
- Stefan M., Bavli S. Recurrent stupor associated with chronic valproic acid therapy and hyperammonemia. Hospital Physician. 2009; 45: 17–20.
- Bajaj J.S., Pinkerton S.D., Sanyal A. J., Heuman D.M. Diagnosis and treatment of minimal hepatic encephalopathy to prevent motor vehicle accidents: a cost-effectiveness analysis. *Hepatology*. 2012 Apr;55(4):1164–71. doi: 10.1002/hep.25507. PMID: 22135042; PMCID: PMC3319334.
- Bogomolov P. O., Bueverov A. O., Uvarova O. V., Matsievich M. V. Hyperammonemia in patients with liver diseases at the pre-cirrotic stage: is it possible? *Clinic. prosp. gastroenterol. Hepatol.* 2013;5:3–8. (In Russ.).
- Hepatic Encephalopathy in Chronic Liver Disease: 2014
 Practice Guideline by the European Association for the
 Study of the Liver and the American Association for
 the Study of Liver Diseases. *Journal of Hepatology*, 2014;
 61(3):642–659. Doi: 10.1016/j.jhep.2014.05.042
- Galvin R., Brathen G., Ivashynka A., Hillbom M., Tanasescu R., Leone M. A. EFNS guidelines for diagnosis, therapy and prevention of Wernicke encephalopathy. European Journal of Neurology. 2010 Dec;17(12):1408–18. doi: 10.1111/j.1468–1331.2010.03153.x. PMID: 20642790.
- Zakharov V. V. Alcohol abuse: neurological complications and modern approaches to therapy. Effective pharmacotherapy. 2014; 8: 36–43. (In Russ.).
- Davis B.C., Bajaj J.S. Effects of Alcohol on the Brain in Cirrhosis: Beyond Hepatic Encephalopathy. *Alcohol Clin Exp Res*. 2018 Apr;42(4):660–667. doi: 10.1111/acer.13605. Epub 2018 Feb 27. PMID: 29417604.
- Huang C. L., Tsai C. J. et al. Alcohol-Related Dementia:
 A Systemic Review of Epidemiological Studies.
 Psychosomatics. 2017 Jul-Aug;58(4):331–342. doi:
 10.1016/j.psym.2017.02.012. Epub 2017 Mar 1. PMID:
 28501289.
- de Oliveira A., Bortolato T., Bernardes Filho F. Pellagra.
 J Emerg Med. 2018 Feb;54(2):238–240. doi: 10.1016/j. jemermed.2017.10.010. Epub 2017 Nov 22. PMID: 29174751.
- Díaz-Fontenla F., Castillo-Pradillo M., Díaz-Gómez A. et al. Refractory hepatic encephalopathy in a patient with hypothyroidism: Another element in ammonia metabolism. World J Gastroenterol. 2017 Jul 28;23(28):5246– 5252. doi: 10.3748/wjg.v23.i28.5246. PMID: 28811719; PMCID: PMC5537191.
- Ermolova T. V., Yakovleva D. M. Efficacy of the use of L-ornithine-L-aspartate in patients with steatohepatitis. Modern gastroenterology and hepatology. 2012, No 1, pp. 22–26.
- Festi D., Vestito A., Mazzell aG., Roda E., Colecchia A. Management of hepatic encephalopathy: focus on antibiotic therapy. *Digestion*. 2006;73 Suppl 1:94–101. doi: 10.1159/000089784. Epub 2006 Feb 8. PMID: 16498257.

- 82. Jalan R., De Chiara F., Balasubramaniyan V., Andreola F., Khetan V., Malago M., Pinzani M., Mookerjee R. P., Rombouts K. Ammonia produces pathological changes in human hepatic stellate cells and is a target for therapy of portal hypertension. *J Hepatol*. 2016 Apr;64(4):823–33. doi: 10.1016/j.jhep.2015.11.019. Epub 2015 Dec 2. PMID: 26654994
- 83. Tomomura M., Tomomura A., Dewan M.A., Saheki T. Long-chain fatty acids suppress the induction of urea cycle enzyme genes by glucocorticoid action. FEBS letters. 1996 Dec 16;399(3):310–2. doi: 10.1016/s0014– 5793(96)01344–0. PMID: 8985169.
- 84. Zhu LH, Armentano LE, Bremmer DR, Grummer RR, Bertics SJ. Plasma concentration of urea, ammonia, glutamine around calving, and the relation of hepatic triglyceride, to plasma ammonia removal and blood acid-base balance. *Journal of dairy science*. 2000 Apr;83(4):734–40. doi: 10.3168/jds.S0022-0302(00)74935-6. PMID: 10791789.
- 85. Jia B, Yu ZJ, Duan ZF, et al. Hyperammonaemia induces hepatic injury with alteration of gene expression profiles. *Liver international*: official journal of the International Association for the Study of the Liver. 2014 May;34(5):748–58. doi: 10.1111/liv.12365. Epub 2013 Nov 20. PMID: 24134218.
- 86. Leung T.M., Lu Y., Yan W., et al. Argininosuccinate synthase conditions the response to acute and chronic ethanol-induced liver injury in mice. *Hepatology*. 2012; 55:1596–1609.
- 87. Yaplito-Lee J., Chow C. W., Boneh A. Histopathological findings in livers of patients with urea cycle disorders. *Mol Genet Metab.* 2013 Mar;108(3):161–5. doi: 10.1016/j. ymgme.2013.01.006. Epub 2013 Jan 23. PMID: 23403242.
- 88. Thomsen K.L., De Chiara F., Rombouts K., Vilstrup H., Andreola F., Mookerjee R. P., Jalan R. Ammonia: A novel target for the treatment of non-alcoholic steatohepatitis. *Med Hypotheses*. 2018 Apr;113:91–97. doi: 10.1016/j. mehy.2018.02.010. Epub 2018 Feb 15. PMID: 29523305.
- Begriche K., Massart J., Robin M.A., Bonnet F., Fromenty B. Mitochondrial adaptations and dysfunctions in nonalcoholic fatty liver disease. *Hepatology*. 2013 Oct;58(4):1497–507. doi: 10.1002/hep.26226. Epub 2013 Aug 7. PMID: 23299992.
- 90. Thomsen K.L., Gronbaek H., Glavind E., et al. Experimental nonalcoholic steatohepatitis compromises ureagenesis, an essential hepatic metabolic function. *Am j physiol Gastrointest liver physiol*. 2014 Aug 1;307(3): G295–301. doi: 10.1152/ajpgi.00036.2014. Epub 2014 Jun 12. PMID: 24924745.
- 91. Thomsen K.L., De Chiara F., Andreola F., et al. Ornithine transcarbamylase gene expression and hepatic urea nitrogen handling are reduced in models of NAFLD and recovers with dietary modulation and reducing bacterial translocation: Rationale for ammonia lowering therapy in NASH patients. Abstract, Annual Meeting of the American Association for the Study of Liver Diseases. 2015.
- 92. Gorban V.V., Korochanskaya N.V., Gorban E.V., Serikova S.N. Clinical-laboratory constellations of non-alcoholic fatty liver disease and the gastroesophageal reflux disease: the possibilities of screening steatohepatosis and steatohepatitis based on the detection of hyperammonemia in outpatient practice. *Experimental and Clinical Gastroenterology.* 2019;(6):105–112. (In Russ.). Doi: 10.31146/1682–8658-ecg-166–6–105–112
- 93. Singh S., Suresh S., McClave S.A., Cave M. Treating Every Needle in the Haystack: Hyperammonemic Encephalopathy and Severe Malnutrition After Bariatric Surgery-A Case Report and Review of the Literature.

- JPENJ Parenter Enteral Nutr. 2015 Nov;39(8):977–85. doi: 10.1177/0148607114546900. Epub 2014 Sep 2. PMID: 25185153.
- 94. Ermolova T.V., Ermolov S. Yu., Belova A. A. Ammonia as a new therapeutic target for chronic liver diseases. *Experimental and Clinical Gastroenterology*. 2020;174(4):24–30. (In Russ.). Doi: 10.31146/1682–8658-ecg-176-4-24-30
- 95. Vaquero J., Chung C., Cahill M. E., Blei A. T. Pathogenesis of hepatic encephalopathy in acute liver failure. *Semin Liver Dis.* 2003 Aug;23(3):259–69. doi: 10.1055/s-2003–42644. PMID: 14523679.
- Uribe M., Moran S., dela M. G. Dietetic manipulations in patients with hepatic encephalopathy. *Rev Gastroenterol Mex*. 1994 Apr-Jun;59(2 Suppl):74–8. (In Spanish). PMID: 7916474.
- Cordoba J., Lopez-Hellin J., Planas M. et al. Normal protein diet for episodic hepatic encephalopathy: results of a randomized study. *J Hepatol*. 2004 Jul;41(1):38–43. doi: 10.1016/j.jhep.2004.03.023. PMID: 15246205.
- Als-Nielsen B., Gluud L. L., Gluud C. Non-absorbable disaccharides for hepatic encephalopathy: systematic review of randomised trials. *BMJ*. 2004 May 1;328(7447):1046. doi: 10.1136/bmj.38048.506134.EE. Epub 2004 Mar 30. PMID: 15054035; PMCID: PMC403844.
- Cash W. J., McConville P., McDermott E. at al. Current concepts in the assessment and treatment of Hepatic Encephalopathy. Q J Med. 2010 Jan;103(1):9–16. doi: 10.1093/qjmed/hcp152. Epub 2009 Nov 10. PMID: 19903725.
- 100. Butterworth R. F. Pathophysiology of hepatic encephalopathy: a new look at ammonia. *Metab Brain Dis.* 2002 Dec;17(4):221–7. doi: 10.1023/a:1021989230535. PMID: 12602499.
- 101. Butterworth R.F., Kircheis G., Hilger N., McPhail M.J.W. Efficacy of L-Ornithine L-Aspartate for the Treatment of Hepatic Encephalopathy and Hyperammonemia in Cirrhosis: Systematic Review and Meta-Analysis of Randomized Controlled Trials. J. Clin. Exp. Hepat. 2018 Sep;8(3):301–313. doi: 10.1016/j.jceh.2018.05.004. Epub 2018 May 22. PMID: 30302048; PMCID: PMC6175748.
- 102. Kizova E.A., Potekhina Yu. P. Effect of l-ornithine-l-aspartate therapy on the hyperammonemia level and results of the number connection test of patients suffering from nonalcoholic fatty liver disease. *Experimental and Clinical Gastroenterology.* 2019; (9):80–84. (In Russ.). Doi: 10.31146/1682–8658-ecg-169–9–80–84
- 103. Demura S, Yamada T, Yamaji S, Komatsu M, Morishita K. The effect of l-ornithine hydrochloride ingestion on performance during incremental exhaustive ergometer bicycle exercise and ammonia metabolism during and after exercise. *Eur J Clin Nutr.* 2010 Oct;64(10):1166–71. doi: 10.1038/ejcn.2010.149. Epub 2010 Aug 18. PMID: 20717126
- 104. Ageeva E. A., Alekseenko S. A. The experience of oral administration of L-ornithine-L-aspartate for hyperammonemia in patients with chronic liver disease at the pre-cirrotic stage. *Clin. Perspective. gastroenterol.* hepatol. 2015, No. 6, pp. 24–26. (In Russ.).
- 105. Burkov S. G., Arutyunov A. G., Godunova S. A. et al. Efficacy of L-Ornithine-L-Aspartate Granules in the Treatment of Non-Alcoholic Fatty Liver Disease. Consilium Medicum. 2010, V. 12, No. 8, pp. 43–47. (In Russ.).
- 106. Osipenko M. F., Redkina A. V., Bikbulatova E. A., et al. Evaluation of L-ornithine-L-aspartate (Hepa-Merz) in the complex treatment of non-alcoholic steatohepatitis.

- Gastroenterology. Supplement to the journal Consilium Medicum. 2010, No. 1, pp. 35–38. (In Russ.).
- 107. Grungreiff K., Lambert-Baumann J. Efficacy of L-Ornithine-L-Aspartate Granules in the Treatment of Chronic Liver Diseases. Suchasna gastroenterology. 2008. No. 2, pp. 59–67. (In Russ.).
- 108. Ermolov S. Yu., Shabrov A. V., Ermolova T. V., et al. New approach to diagnostics and correction of portohepatic hemodynamics in patients with chronic liver diseases. *Eksp Klin Gastroenterol*. 2007;(4):82–7. (In Russ.).
- 109. Gromova O. A., Torshin I. Yu., Maximov V. A., Gromov A. N., Rudakov K. V. Systematic analysis of lactitol studies. Experimental and Clinical Gastroenterology. 2019;162(2): 131–142. (In Russ.). DOI: 10.31146/1682– 8658-ecg-162-2-131-142
- 110. Evdokimova A.G., Tomova A.V., Tereshchenko O.I., Zhukolenko L.V., Evdokimov V.V. Clinical efficacy of the receptor beta and alpha blockers – carvedilol and ornithine-aspartate in patients with CHD and CHF combined with alcoholic liver disease. *Experimental* and Clinical Gastroenterology. 2016;(6):42–47. (In Russ.).
- Bajaj J. S., Riggio O. Drug therapy: Rifaximin. Hepatology. 2010 Oct;52(4):1484–8. doi: 10.1002/hep.23866. PMID: 20814894.
- 112. Kimer N., Krag A., Gluud L. L. Safety, efficacy, and patient acceptability of Rifaximin for hepatic encephalopathy. *Patient Prefer. Adherence*. 2014 Mar 18;8:331–8. doi: 10.2147/PPA.S41565. PMID: 24672227; PMCID: PMC3964161.
- 113. Sharma B. C., Sharma P., Lunia M. K., Srivastava S., Goyal R., Sarin S. K. A randomized, double-blind, controlled trial comparing Rifaximin plus lactulose with lactulose alone in treatment of overt hepatic encephalopathy. Am. J. Gastroenterol. 2013 Sep;108(9):1458–63. doi: 10.1038/ajg.2013.219. Epub 2013 Jul 23. PMID: 23877348.
- 114. Loguercio C., Federico A., De Girolamo V., Ferrieri A., Del Vecchio B. C. Cyclic treatment of chronic hepatic encephalopathy with rifaximin. Results of a doubleblind clinical study. *Minerva Gastroenterol Dietol*. 2003 Mar;49(1):53–62. PMID: 16481971.
- 115. Williams R., James O. F., Warnes T. W., Morgan M. Y. Evaluation of the efficacy and safety of rifaximin in the treatment of hepatic encephalopathy: a double-blind, randomized, dose-finding multi-centre study. Eur J Gastroenterol Hepatol. 2000 Feb;12(2):203–8. doi: 10.1097/00042737–200012020–00012. PMID: 10741936.
- 116. Riordan S.M., Williams R. Treatment of hepatic encephalopathy. N Engl J Med. 1997 Aug 14;337(7):473–9. doi: 10.1056/NEJM199708143370707. PMID: 9250851.
- 117. Patidar K.R., Thacker L.R., Wade J.B., et al. Covert hepatic encephalopathy is independently associated with poor survival and increased risk of hospitalization. *Am J Gastroenterol*. 2014 Nov;109(11):1757–63. doi: 10.1038/ajg.2014.264. Epub 2014 Sep 2. PMID: 25178701; PMCID: PMC4321782.
- 118. Ivashkin V. T., Mayevskaya M. V., Pavlov C. S., Fedosyina Y. A., Bessonova Y. N., Pirogova I. Y., Garbuzenko D. V. Treatment of liver cirrhosis complications: Clinical guidelines of the Russian Scientific Liver Society and Russian gastroenterological association. Russian Journal of Gastroenterology, Hepatology, Coloproctology. 2016;26(4):71–102. (In Russ.). Doi: 10.22416/1382–4376–2016–26–4–71–102
- 119. Festi D. et al. Rifaximin in the treatment of chronic hepatic encephalopathy; results of a multicenter study of efficacy and safety. Curr Ther Res. 1993; 54(5): 598–609

- 120. Dhiman R.K., Thumburu K.K., Verma N., Chopra M., Rathi S., Dutta U., Singal A.K., Taneja S., Duseja A., Singh M.; Indian National Association for Study of Liver (INASL) Hepatic Encephalopathy Study Group (IHESG). Comparative Efficacy of Treatment Options for Minimal Hepatic Encephalopathy: A Systematic Review & Network Meta-analysis. Clin Gastroenterol Hepatol. 2019 Aug 30. pii: S1542–3565(19)30969–3. doi: 10.1016/j.cgh.2019.08.047
- 121. Bajaj J.S., Saeian K., Christensen K.M., et al. Probiotic yogurt for the treatment of minimal hepatic encephalopathy. Am J Gastroenterol. 2008 Jul;103(7):1707–15. doi: 10.1111/j.1572-0241.2008.01861.x. PMID: 18691193.
- 122. Sharma P., Sharma B. C., Puri V., Sarin S. K. An open-label randomized controlled trial of lactulose and probiotics in the treatment of minimal hepatic encephalopathy. *Eur J Gastroenterol Hepatol*. 2008 Jun;20(6):506–11. doi: 10.1097/MEG.0b013e3282f3e6f5. PMID: 18467909.
- 123. Bongaerts G., Severijnen R., Timmerman H. Effect of antibiotics, prebiotics and probiotics in treatment for hepatic encephalopathy. *Med. Hypotheses*. 2005;64(1):64–8. doi: 10.1016/j.mehy.2004.07.029. PMID: 15533613.
- 124. Dalal R., McGee R.G., Riordan S. M., Webster A. C. Probiotics for people with hepatic encephalopathy. Cochrane Database Syst Rev. 2017 Feb 23;2(2): CD008716. doi: 10.1002/14651858.CD008716.pub3. PMID: 28230908; PMCID: PMC6464663.
- 125. Bajaj D., Chahal P. A case of multiple misdiagnoses in a septuagenarian. *Am. J. Med.* 2014 Jul;127(7): e15-6. doi: 10.1016/j.amjmed.2014.01.022. Epub 2014 Apr 19. PMID: 24751123.

- 126. Macbeth W.A., Kass E., McDermott W. Treatment of hepatic encephalopathy by alteration of intestinal flora with Lactobacillus acidophilus. *Lancet.* 1965 Feb 20;-1(7382):399–403. doi: 10.1016/s0140–6736(65)90002–4. PMID: 14238091.
- 127. Loguercio C., Del Vecchio Blanco C., Coltorti M. Enterococcus lactic acid bacteria strain SF68 and lactulose in hepatic encephalopathy: A controlled study. J. Int. Med. Res. 1987 Nov-Dec;15(6):335–43. doi: 10.1177/030006058701500602. PMID: 3125077.
- 128. Dhiman R.K., Rana B., Agrawal S., et al. Probiotic VSL# 3 reduces liver disease severity and hospitalization in patients with cirrhosis: a randomized, controlled trial. *Gastroenterology*. 2014 Dec;147(6):1327–37.e3. doi: 10.1053/j.gastro.2014.08.031. Epub 2014 Aug 27. PMID: 25450083.
- 129. Ilchenko L. Yu. Hepatic encephalopathy. Collection of reasearch works: Selected chapters of clinical gastroenterology. Moscow. Anaharsis Publ., 2005., pp. 209–18. (In Russ.).
- 130. Lazebnik L. B., Tarasova L. V., Komarova E. A., Tsyganova Yu. V., Busalaeva E. I. Changes in the concentration of ammonia and other biochemical parameters in patients with new coronavirus infection. *Experimental and Clinical Gastroenterology*. 2021. (4), (In Russ.). (in press)
- 131. Grinevich V.B., Kravchuk Yu.A., Tkachenko E. I., Pershko A. M., Ped V. I., Sas I. E., Gubonina I. V., Lazebnik L. B., Stefanyuk O. V. Features of management of patients with gastroenterological pathology in the conditions of the COVID-19 pandemic. *Experimental and Clinical Gastroenterology*. 2020;174(4):3–18. (In Russ.). Doi: 10.31146/1682–8658-ecg-176–4–3–18